



Thaw and Culture Details

Cell Line Name	JHU250i
WiCell Lot Number	DB36904
Provider	Johns Hopkins University – Laboratory of Dr. Lewis Becker
Banked By	Johns Hopkins University – Laboratory of Dr. Lewis Becker
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 4 wells of a 6 well plate using TeSR™-E8™ and Recombinant Human Vitronectin. WiCell recommends thawing using ROCK Inhibitor for best results.
Protocol	WiCell Feeder Independent Pluripotent Stem Cell Protocol
Culture Platform Prior to Freeze	Feeder Independent
	Medium: E8
	Matrix: Vitronectin
Passage Number	p7 These cells were cultured for 7 passages post reprogramming prior to freeze. Add +1 to the passage number to best represent the overall passage number of the cells at thaw.
Date Vialied	03-September-2015
Vial Label	P250 P7 1.5x10 ⁶ 9/3/15
Biosafety and Use Information	This cell line is of human origin. Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.

Testing Performed by WiCell

Test Description	Test Provider	Test Method	Test Specification	Result
Karyotype by G-banding	WiCell	SOP-49	Expected karyotype	See Report
Post-Thaw Viable Cell Recovery	WiCell	SOP-99	Recoverable attachment after passage	Pass
Identity by STR	WiCell	PowerPlex 16 HS System by Promega	Defines STR profile of deposited cell line	Pass
Sterility	Steris	ST/07	Negative	Pass
Mycoplasma	WiCell	SOP-79	Negative	Pass

Testing Reported by Provider

The Provider stated that some or all of the additional analyses listed below may have been performed for this cell line. For more information, publication and dbGaP links, where available, are provided on the cell line specific web page on the WiCell website.

- Embryoid bodies
- Infinium® Expanded Multi-Ethnic Genotyping Array (MEGA^{EX})



Approval Date	Quality Assurance Approval
14-July-2016	<p style="text-align: right;">12/9/2020</p> <p>X JKG _____ WiCell Quality Assurance Signed by: Gay, Jenna</p>

Date Reported: Tuesday, November 17, 2020

Cell Line Sex: Male

Cell Line: JHU250i-DB36904

Reason for Testing: LOT_RELEASE

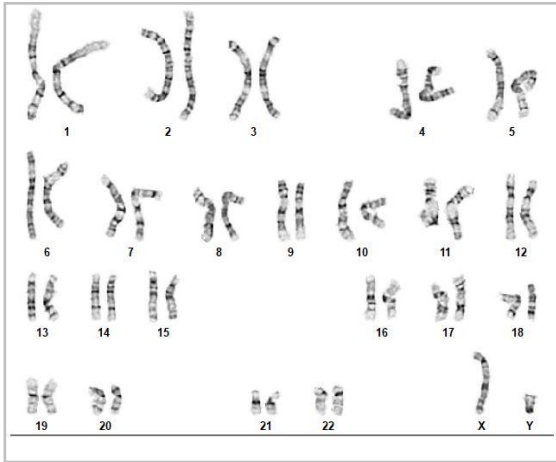
Submitted Passage #: 8

Date of Sample: 11/6/2020

Investigator: WiCell Stem Cell Bank, WiCell

Specimen: Human iPSC

Results: 46,XY



Cell: 63

Slide: G03

Slide Type: Karyotype

Total Counted: 20

Total Analyzed: 8

Total Karyogrammed: 4

Band Resolution: 475 - 550

Interpretation:

This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.

Completed by: [Redacted]

Reviewed and Interpreted by: [Redacted], PhD, FACMG

Date: _____ **Sent By:** _____ **Sent To:** _____ **QC Review By:** _____

Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e., mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

This assay was conducted solely for listed investigator/institution. The results of this assay are for research use only. Unless otherwise mutually agreed in writing, the services provided to you hereunder by WiCell Research Institute, Inc. ("WiCell") are governed solely by WiCell's Terms and Conditions of Service, found at www.wicell.org/privacyandterms. Any terms you may attach to a purchase order or other document that are inconsistent, add to, or conflict with WiCell's Terms and Conditions of Service are null and void and of no legal force or effect.



Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell
 Samples Received: 02Nov20, 06Nov20
 STR Amplification Date: 11Nov20

Sample Name	WA09-RB67589 p.30	JHU105i- DB36241 p.5	JHU148i-DB36280 p.7	JHU102i-DB41279 p.5	JHU250i- DB36904 p.8	JHU173i-DB36380 p.10
Label on tube	83593	83677	83678	83679	83687	83688
FGA	Identifying information has been redacted to protect donor confidentiality. If more information is required, please contact info@wicell.org					
TPOX						
D8S1179						
vWA						
Amelogenin						
Penta_D						
CSF1PO						
D16S539						
D7S820						
D13S317						
D5S818						
Penta_E						
D18S51						
D21S11						
TH01						
D3S1358						
Allelic Polymorphisms	24	30	27	25	25	43
Matches*	See Matches Comment					
Comments						See Mixed Cell Line Comment

**Note: The STR profile of the following sample is an exact match for the given sample/samples.*



Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell
Samples Received: 02Nov20, 06Nov20
STR Amplification Date: 11Nov20

Results: The genotypic profiles comprise a range of 24-43 allelic polymorphisms across the 15 STR loci analyzed.

Interpretation: The concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. These results suggests that the cells submitted correspond to the cell lines as named and were not contaminated with any other human cells or a significant amount of mouse feeder layer cells.

Sensitivity: Sensitivity limits for detection of STR polymorphisms unique to either this or other human cell lines is ~2-5%.

Mixed Cell Line: Sample 83688 shows signs of possible contamination. The most likely explanation for this result is that two cultures have been mixed. G-banded karyotype results confirmed the presence of both male and female cells in the culture. Please resubmit this sample.

Matches: Sample 83593 is an exact match to 14630, 74319, 74844, 74924, and 74925.

12/1/2020

X [Redacted]

Tech #1

Characterization

Signed by: [Redacted]

12/1/2020

X [Redacted]

Tech #2

Characterization

Signed by: [Redacted]

12/1/2020

X [Redacted]

QA Review

Quality Assurance

Signed by: [Redacted]

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Raw data is available upon request.

Native Product Sterility Report



WiCell
504 S Rosa Rd, Rm 101
Madison, WI 53719

SAMPLE #: 18050738
DATE RECEIVED: 10-May-18
TEST INITIATED: 14-May-18
TEST COMPLETED: 29-May-18

SAMPLE NAME / DESCRIPTION: UCSD165i-97-1 WB66795 13679
UCSD224i-NDC1-2 WB66797 13680
UCSD224i-NDC1-2 WB66798 13681
UWWC1-DS4 WB66799 13682
WC035i-SOD1-D90D WB66757 13683
JHU018i DB40957 13684
JHU032i DB36206 13685
JHU083i DB41146 13686
JHU126i DB36258 13687
JHU167i DB41380 13688
JHU190i DB36770 13689
JHU240i DB41420 13690
JHU054i DB41080 13691
JHU188i DB36766 13692
JHU084i DB41149 13693
JHU224i DB36895 13694
JHU250i DB36904 13695
JHU221i DB36885 13696
JHU218i DB36874 13697
JHU217i DB36868 13698

UNIQUE IDENTIFIER: NA
PRODUCT REGISTRATION: Other: Human iPS cells

TEST RESULTS:

# Tested	# Positives (Growth)	- Control
20	0	3 Negatives

TEST SUMMARY:

# Samples	Media Type	Volume (mL)	Incubation Temperature (° C)	Incubation Duration (Days)
20	TSB	40	20-25	14
20	FTG	40	30-35	14

Native Product Sterility Report



REFERENCE: Processed according to LAB-003: Sterility Test Procedure
METHOD VALIDATION / PD #: 000053
TEST METHODOLOGY: USP - Direct Transfer

COMMENTS: Sample #18050738

REVIEWED BY

A handwritten signature in blue ink, consisting of a large, stylized 'S' followed by a horizontal line.

DATE

30 MAY 18

Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. This test report shall not be reproduced, except in full, without prior written approval. Liability is limited to the costs of the tests.



Mycoplasma Assay Report

PCR-based assay performed by WiCell

WiCell

04Nov20

FORM SOP-83.01

Version 01

Sample Name	Result	Comments/Suggestions
INC149 02Nov20AP (83598)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
INC123 02Nov29KR 1 of 2 (83599)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
INC123 02Nov20KR 2 of 2 (83600)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
INC169 02Nov20MMM 1 of 2 (83601)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
INC169 02Nov20MMM 2 of 2 (83602)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU105i-DB36241 (83622)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU004i-2-DB40945 (83623)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU036i-DB40981 (83624)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU102i-DB41279 (83625)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU148i-DB36280 (83626)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU173i-DB36380 (83627)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU214i-DB36851 (83628)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU234i-DB37041 (83629)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
JHU250i-DB36904 (83630)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
Positive (+) Control	Positive	
Negative (-) Control	Negative	

Reported by: [REDACTED], Senior Cell Culture Specialist

Reviewed by: [REDACTED], Assistant Cell Culture Specialist

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A gel image is available upon request.